

## Mouse Tail Snip Procedure

<b>Equipment:</b> Hot Bead Sterilizer	Cotton Balls
70% Ethyl Alcohol	3-4 Paper Towels
Sharp-Blunt Scissors	Isoflurane Anesthetic
1 Pr. Nitrile Gloves	Ear Punch
Anesthetic Jar	Ruler
Ear tags & applier	250 ml Glass beaker
Gauze sponges	DNA away™ / Eliminate™

**Preparation:** Before starting the tail snip procedure, REMOVE the lid and turn on the Hot Bead Sterilizer. Allow the temperature to reach 250°C. While the sterilizer is warming up, prepare the mice by sexing and weaning from the nursery cage. Determine the ID numbers for the mice and record on their cage cards and on the tail snip vials. Record the procedure in the Laboratory Notebook for that group- include the date/day, parents, generation, ID# with sex and coat color. Place the stacked paper towels on the counter to prevent loss of body heat from the anesthetized mice.

**Procedure:** Wearing nitrile gloves, charge the anesthetic chamber using Isoflurane anesthetic. Under the hood, soak one cotton ball with the anesthetic and place in the jar under the platform and plastic mesh. Ear punch the 1<sup>st</sup> mouse using the “Pollard Ear Punch Code”, tattoo the tail, or place an ear tag (all CTR mice) and place into the anesthetic jar. Place the scissors in the sterilizer. Do Not insert the scissor blades too deeply into the beads to avoid over heating the handles. Remove the anesthetized mouse from the jar and place on the paper towels. Grasp the tail securely. Remove the scissors from the sterilizer and cut a 1- 2 cm length from the end of the tail. Cauterize the tail using the flat of the scissor blade. Check for any hemorrhage. Replace the mouse in the cage. The tail snip is placed in the labeled vial. The scissor blades are wiped with an alcohol soaked gauze sponge to remove any tissue and blood. Prior to returning them to the sterilizer the scissor blades are soaked in DNA away™ or Eliminate™ in a 250 ml glass beaker. This procedure is repeated until all tail snips have been collected. Double check the mice for bleeding before returning the cages to the animal room.

**Clean-up:** The sterilizer is turned off and the lid replaced. The instruments and equipment are washed in Nolvasan Surgical Scrub. The anesthetic jar lid is removed and allowed to air under the hood until all anesthetic vapors have evaporated. The used DNA away™ or Eliminate™ is discarded and the beaker washed. Instrument scrub brushes are in the drawer with the instruments. The ear punch is washed then rinsed with alcohol. The ear punch MUST be towel dried to prevent rusting. The instruments are replaced in the instrument drawer when dry. The counter top is cleaned using the disinfectant spray provided in the room. All paper waste is placed in the room trash can ( autoclave bag liner).