

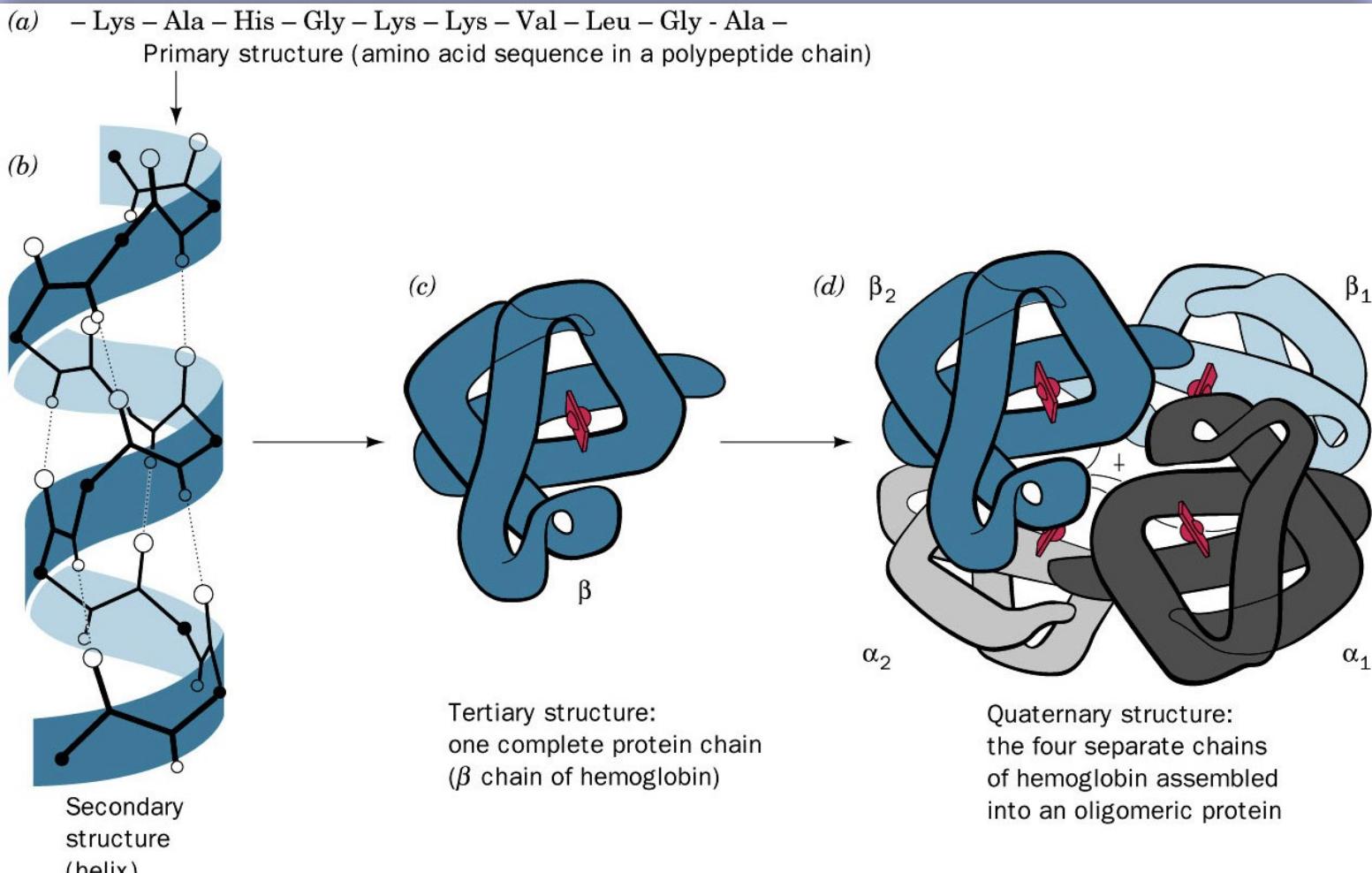
## **Proteins Primary Structure**

### **Peptide/Protein Sequencing; Chemical Synthesis**

**CHEM 420 – Principles of Biochemistry**  
**Instructor – Anthony S. Serianni**

**Chapter 7: Voet/Voet, *Biochemistry*, 2011**  
**Fall 2015**

**September 7 & 9**

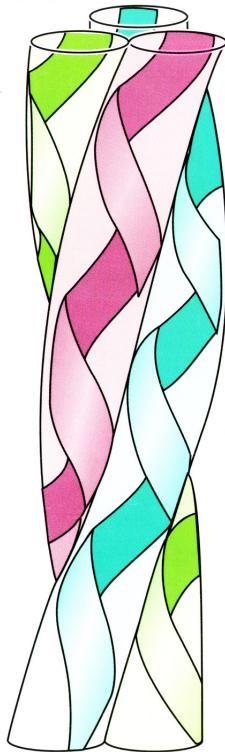


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## Structural hierarchy of proteins

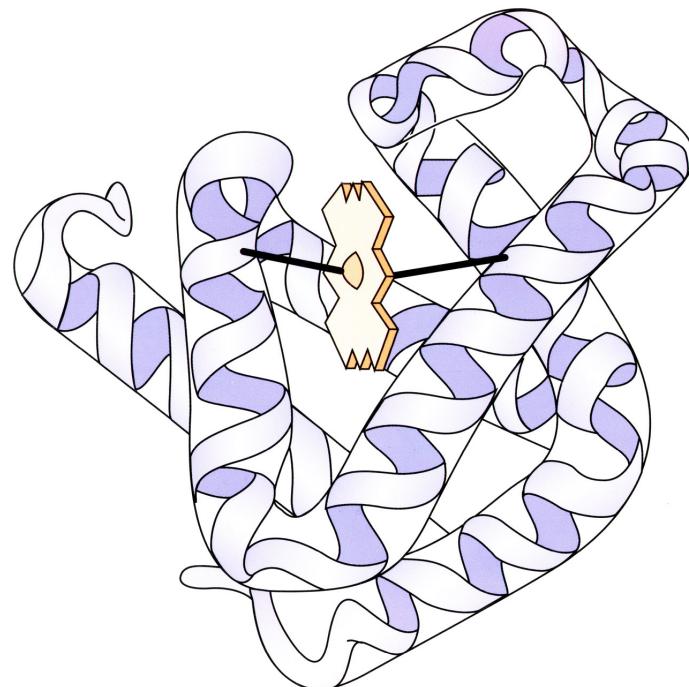
## Representations of fibrous and globular proteins

(a)

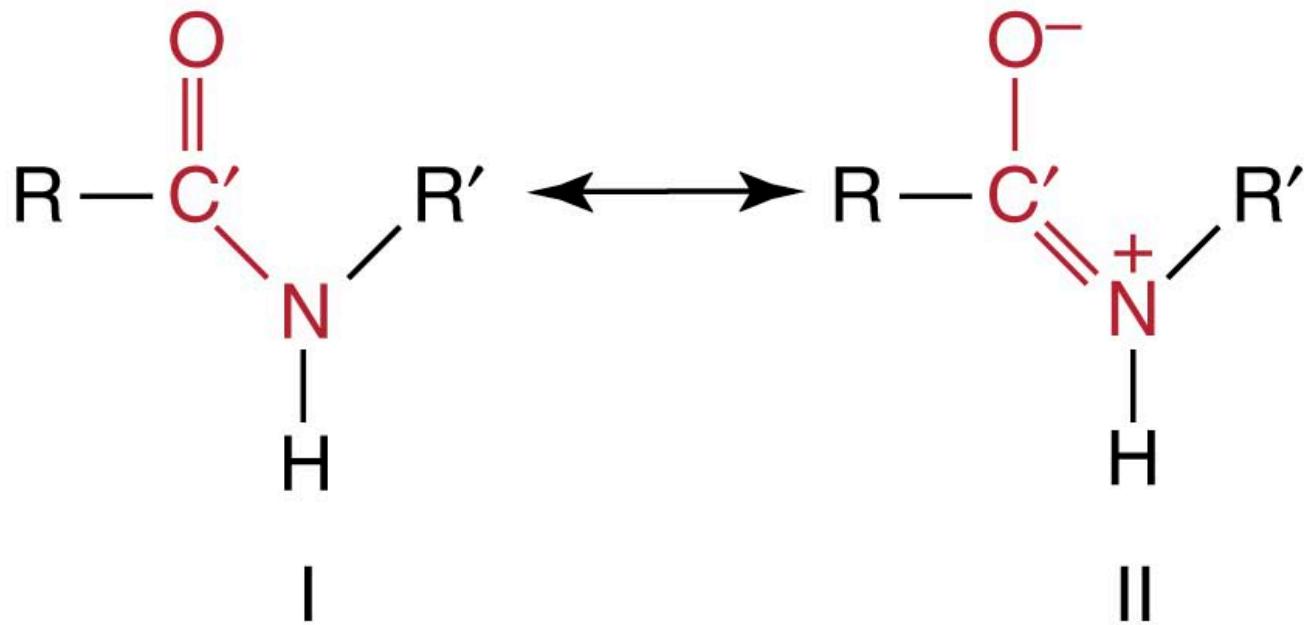


**Collagen, a  
fibrous protein**

(b)



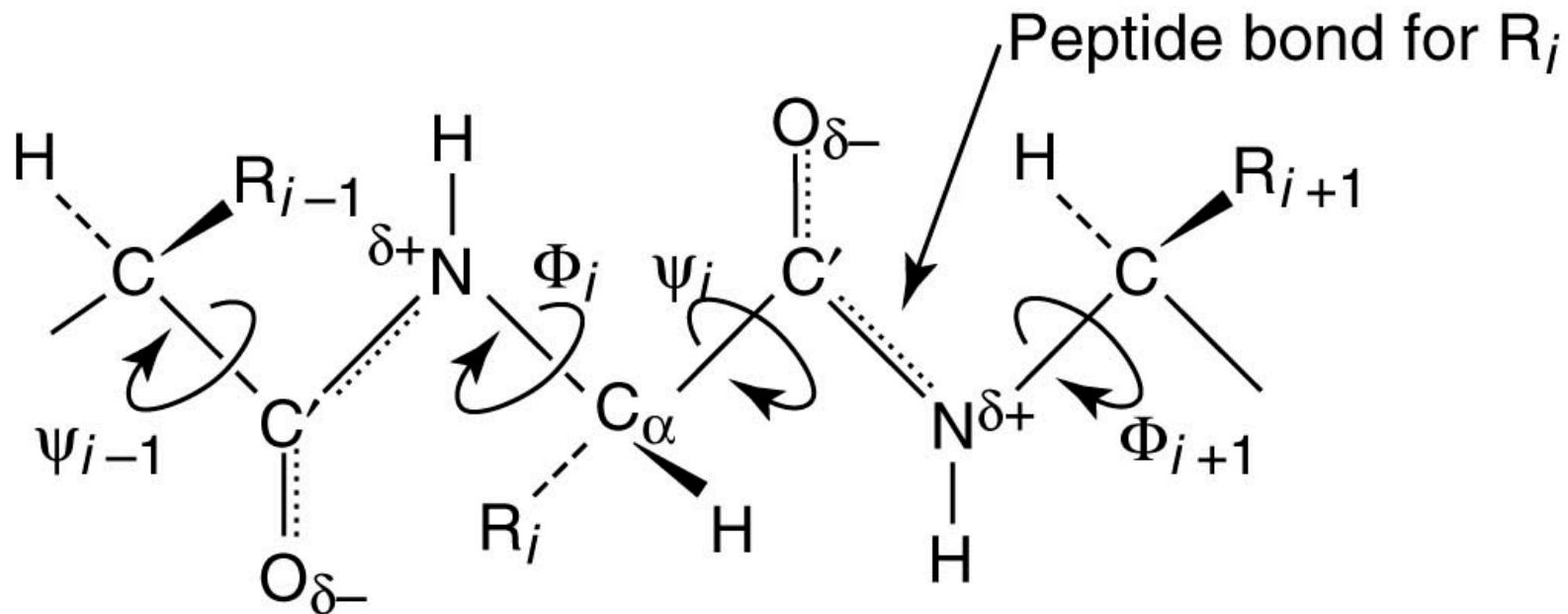
**Myoglobin, a globular protein**



**Figure 3.10. Electronic isomer structures of a peptide bond.**

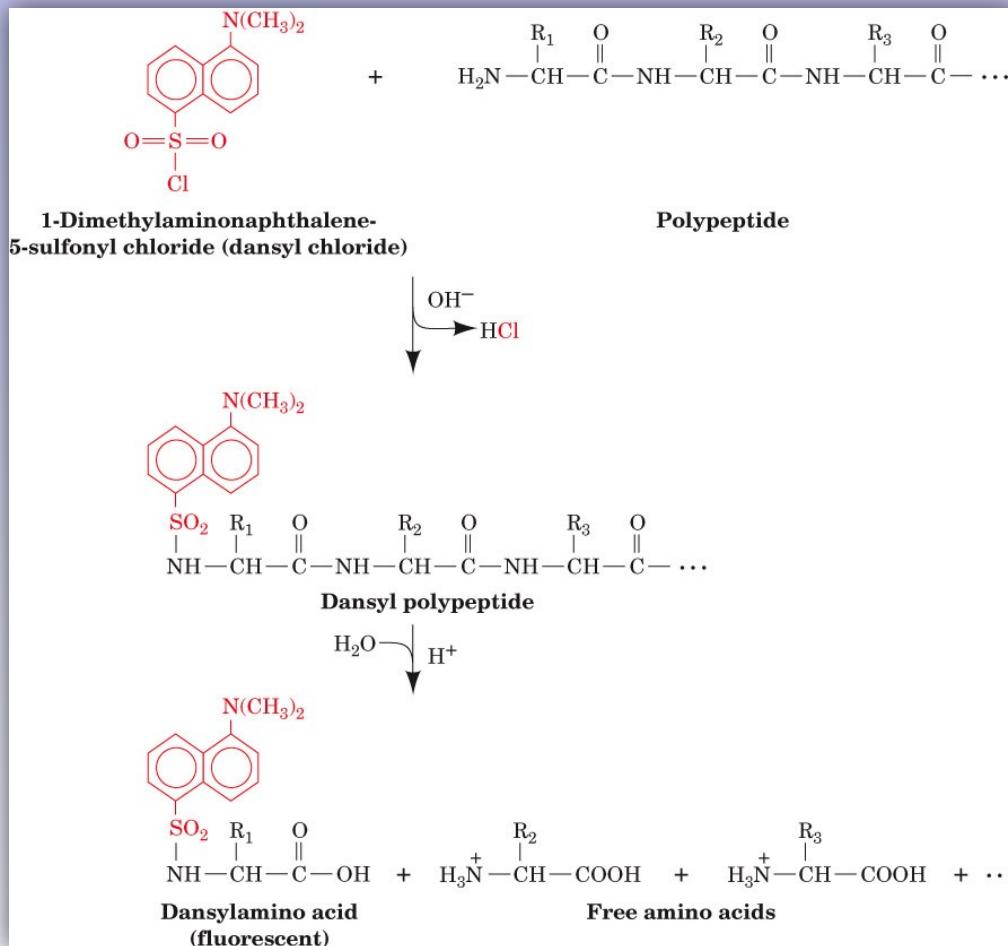
*Textbook of Biochemistry With Clinical Correlations, Sixth Edition, Edited by Thomas M. Devlin. Copyright © 2006 John Wiley & Sons, Inc.*

## $\phi$ , $\psi$ and peptide bonds along the backbone of a polypeptide

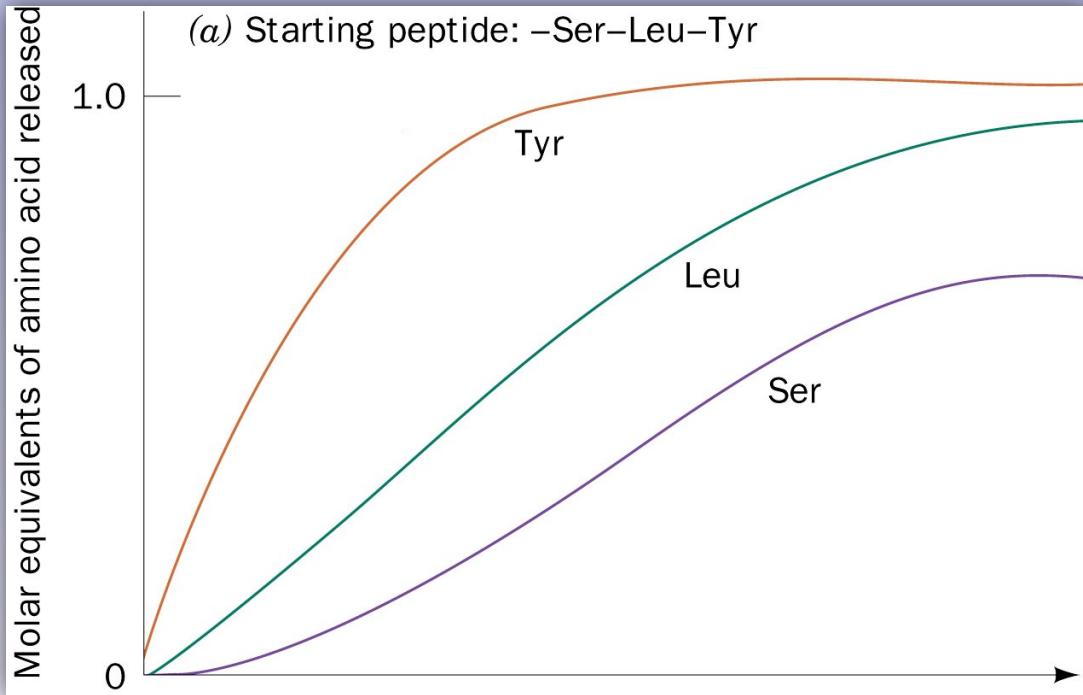


**Figure 3.11. Amino acid residue within a polypeptide chain.**

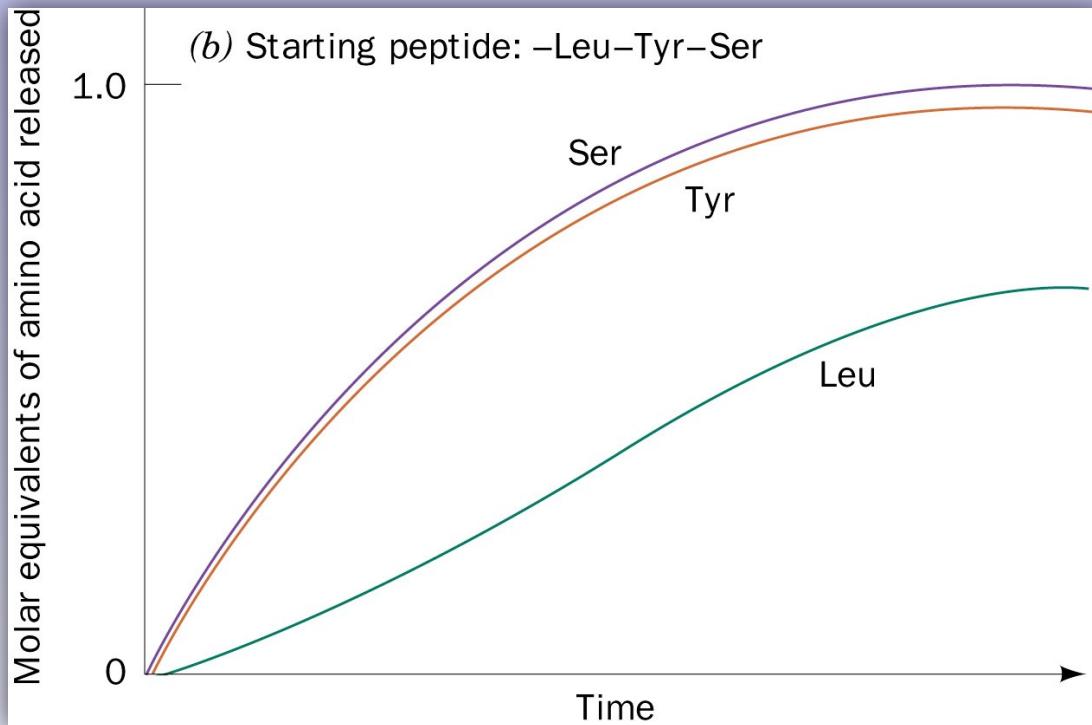
# Determining the 1° structure (sequence) of a polypeptide



The reaction of dansyl chloride in end-group (N-terminus) analysis of a protein



Hypothetical rates of carboxypeptidase-catalyzed  
release of amino acids from the C-terminus of a protein:  
**all bonds cleaved at the same rate**

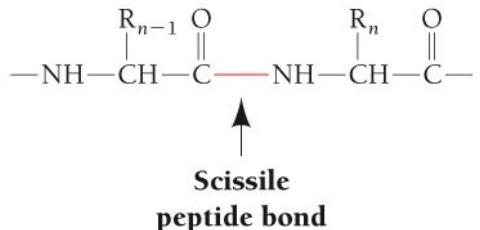


Hypothetical rates of carboxypeptidase-catalyzed release of amino acids from the C-terminus of a protein: **Ser slow, Tyr fast, and Leu intermediate**

Enzyme	Source	Specificity <sup>a</sup>
Carboxypeptidase A	Bovine pancreas	$R_n \neq Arg, Lys, Pro; R_{n-1} \neq Pro$
Carboxypeptidase B	Bovine pancreas	$R_n = Arg, Lys; R_{n-1} \neq Pro$
Carboxypeptidase C	Citrus leaves	All free C-terminal residues; pH optimum = 3.5
Carboxypeptidase Y	Yeast	All free C-terminal residues, but slowly with $R_n = Gly$
Leucine aminopeptidase	Porcine kidney	$R_1 \neq Pro$
Aminopeptidase M	Porcine kidney	All free N-terminal residues

<sup>a</sup> $R_1$  = the N-terminal residue;  $R_n$  = the C-terminal residue.

## Specificities of various **exopeptidases**: C-terminus and N-terminus

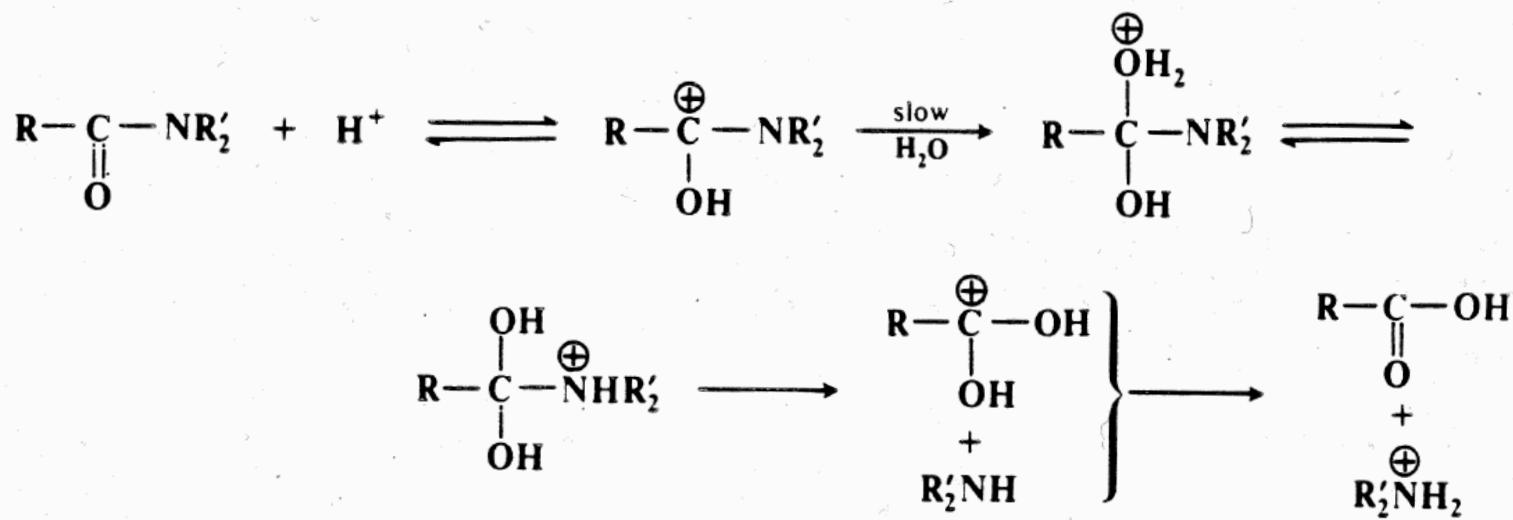
Enzyme	Source	Specificity	Comments
		 <b>Scissile peptide bond</b>	
Trypsin	Bovine pancreas	$R_{n-1} = \text{positively charged residues: Arg, Lys}; R_n \neq \text{Pro}$	Highly specific
Chymotrypsin	Bovine pancreas	$R_{n-1} = \text{bulky hydrophobic residues: Phe, Trp, Tyr}; R_n \neq \text{Pro}$	Cleaves more slowly for $R_{n-1} = \text{Asn, His, Met, Leu}$
Elastase	Bovine pancreas	$R_{n-1} = \text{small neutral residues: Ala, Gly, Ser, Val}; R_n = \text{Pro}$	
Thermolysin	<i>Bacillus thermoproteolyticus</i>	$R_n = \text{Ile, Met, Phe, Trp, Tyr, Val}; R_{n-1} \neq \text{Pro}$	Occasionally cleaves at $R_n = \text{Ala, Asp, His, Thr}$ ; heat stable
Pepsin	Bovine gastric mucosa	$R_n = \text{Leu, Phe, Trp, Tyr}; R_{n-1} \neq \text{Pro}$	Also others; quite nonspecific; pH optimum 2
Endopeptidase Arg-C	Mouse submaxillary gland	$R_{n-1} = \text{Arg}$	May cleave at $R_{n-1} = \text{Lys}$
Endopeptidase Asp-N	<i>Pseudomonas fragi</i>	$R_n = \text{Asp}$	May cleave at $R_n = \text{Glu}$
Endopeptidase Glu-C	<i>Staphylococcus aureus</i>	$R_{n-1} = \text{Glu}$	May cleave at $R_{n-1} = \text{Gly}$
Endopeptidase Lys-C	<i>Lysobacter enzymogenes</i>	$R_{n-1} = \text{Lys}$	May cleave at $R_{n-1} = \text{Asn}$

**Specificities of various endopeptidases:  $R_{n-1}$  and  $R_n$  recognition**

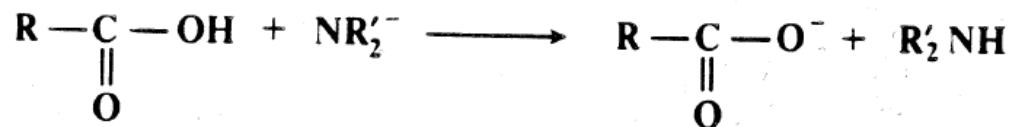
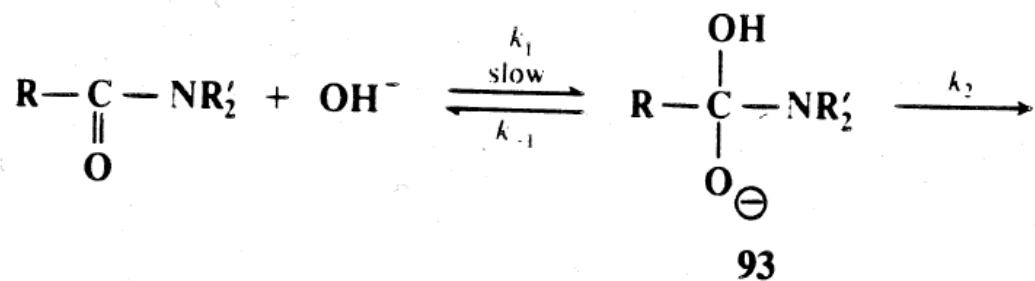
**Enzyme mechanism: Serine proteases**

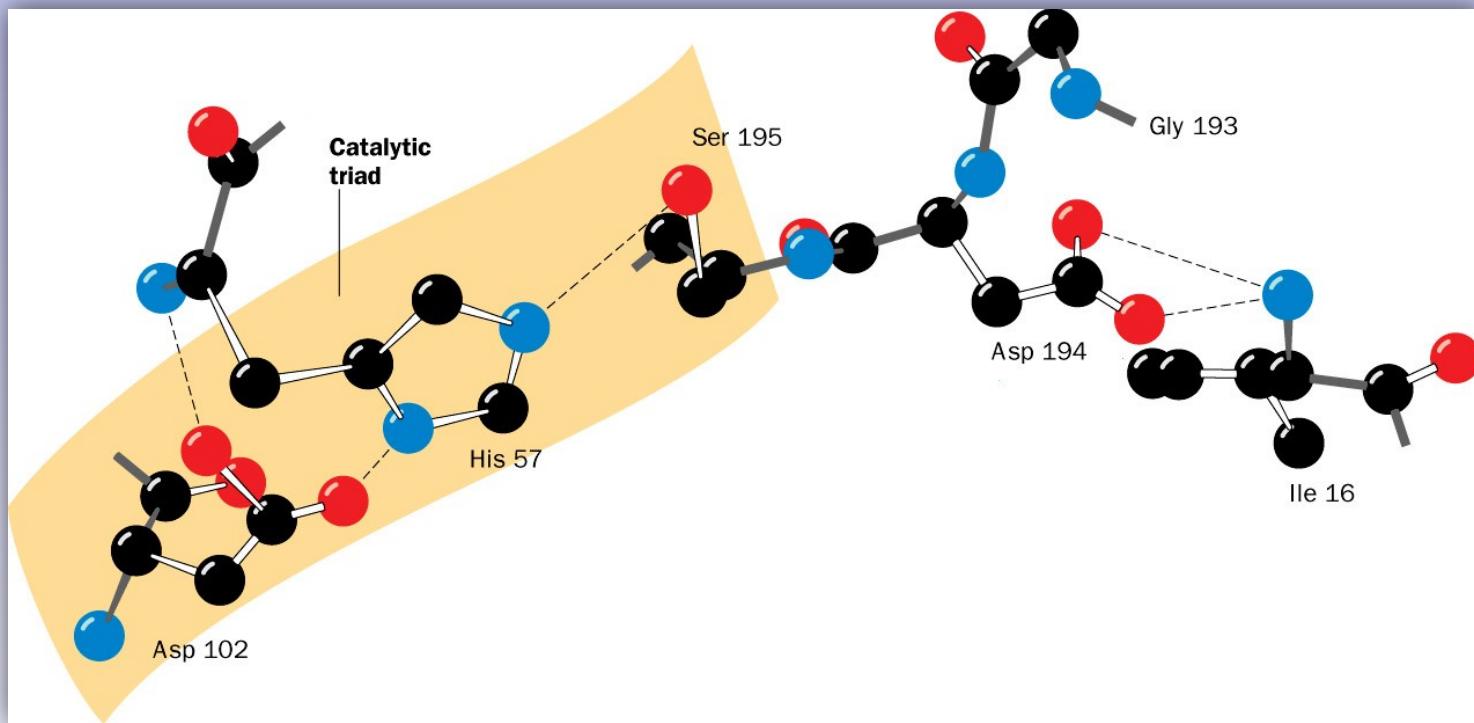
**Trypsin and Chymotrypsin**

## Amide Bond Hydrolysis: Acid-catalyzed Mechanism



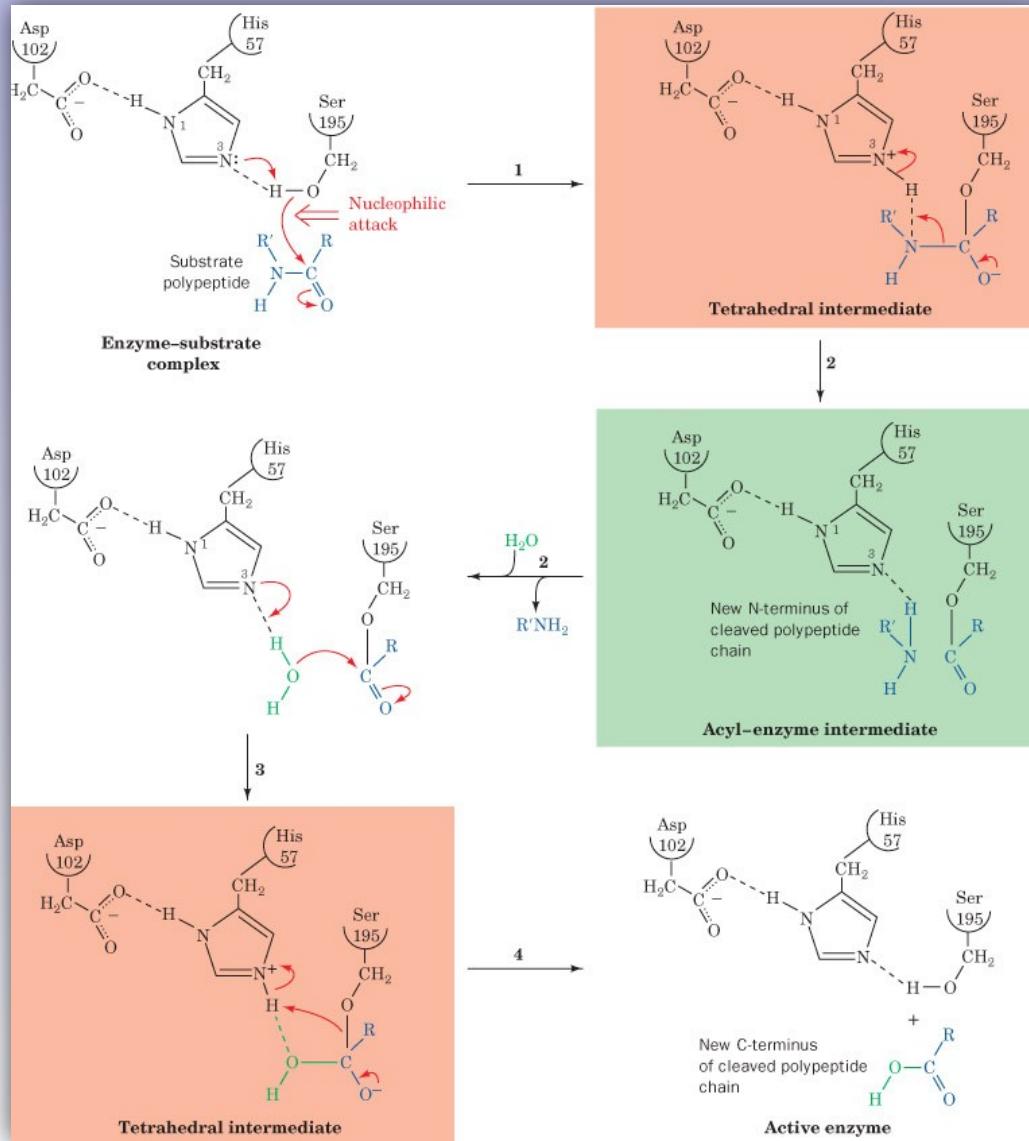
## Amide Bond Hydrolysis: Base-catalyzed Mechanism



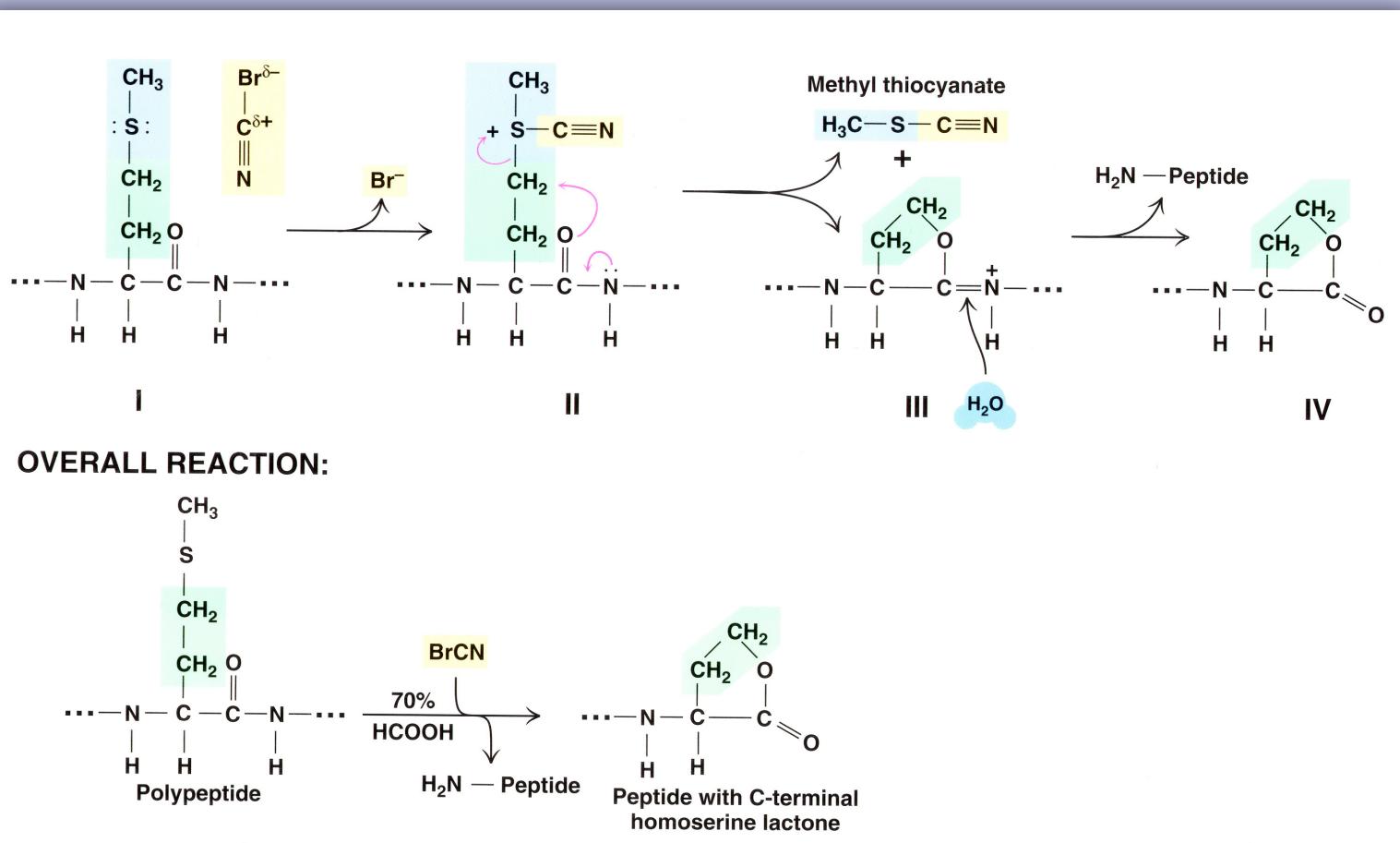


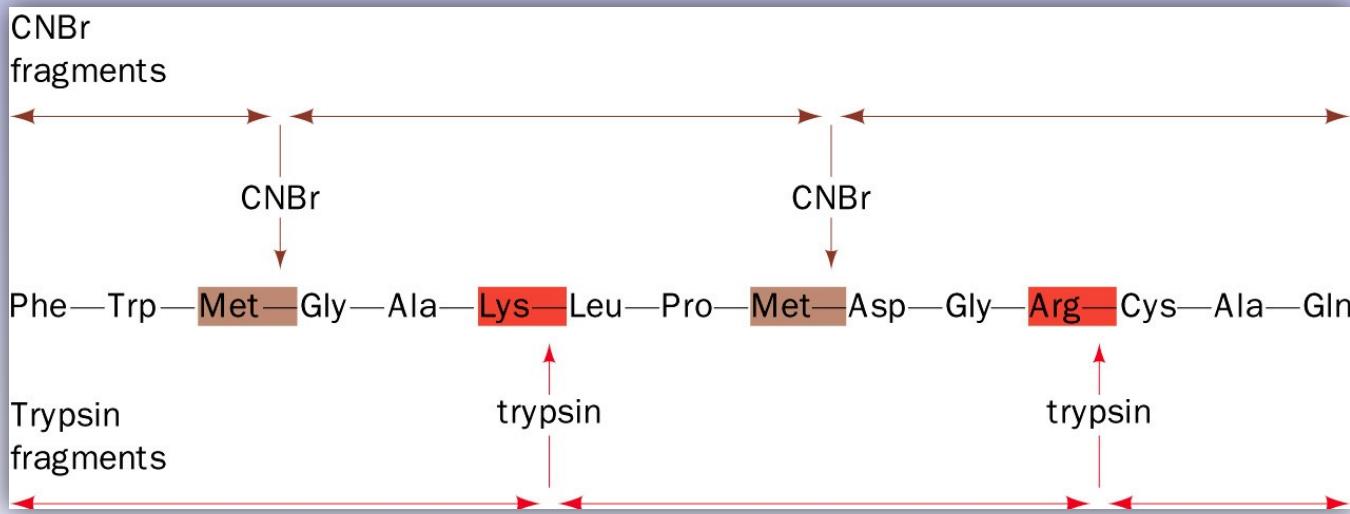
Active site residues of chymotrypsin  
(catalytic triad): a serine protease

# Catalytic mechanism of serine proteases

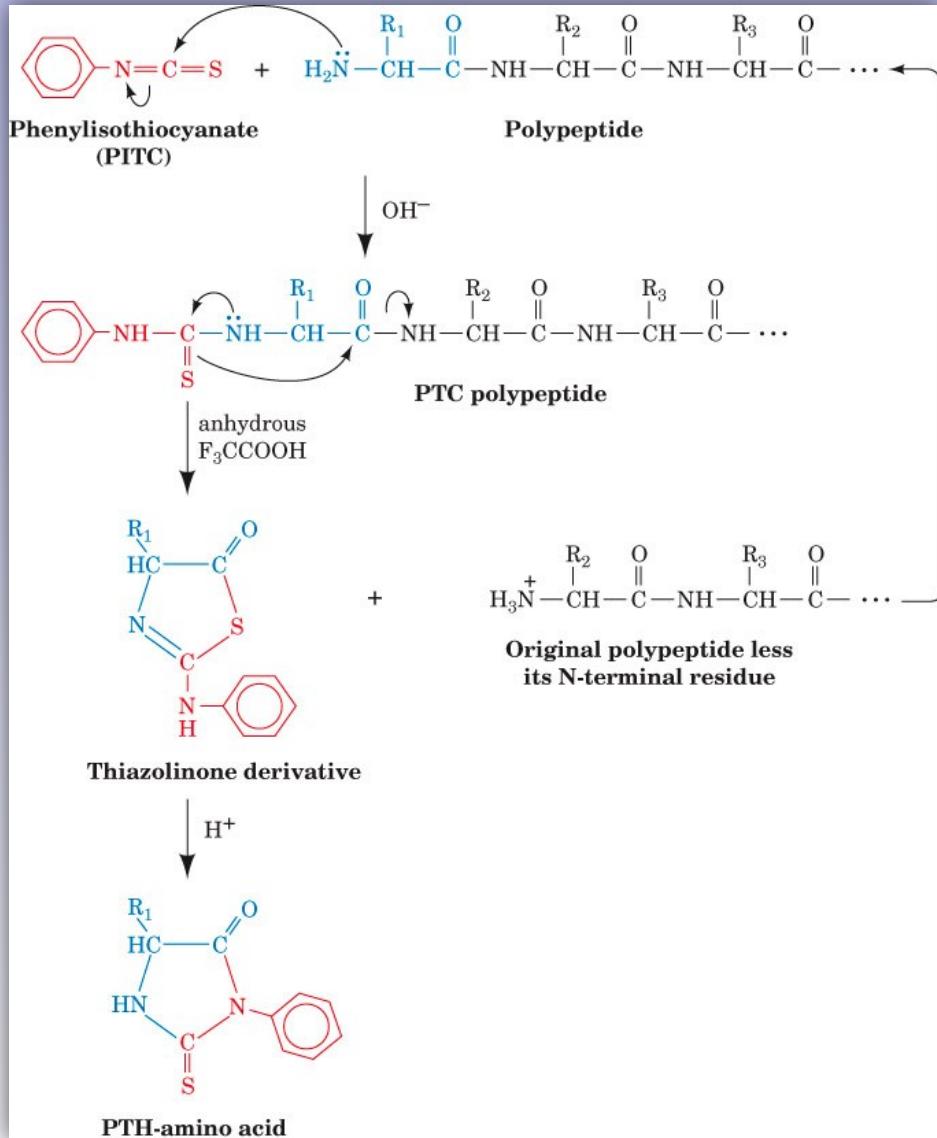


# Cyanogen bromide mediated cleavage of a peptide bond



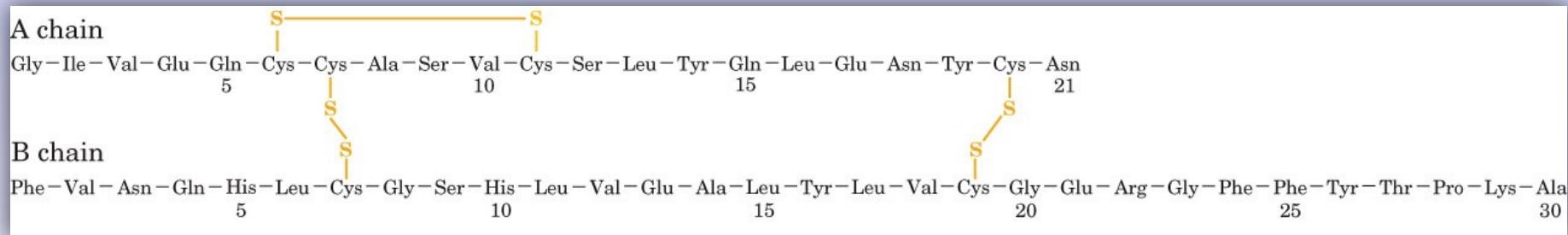


The amino acid sequence of an oligopeptide showing its susceptibility to CNBr and trypsin cleavage



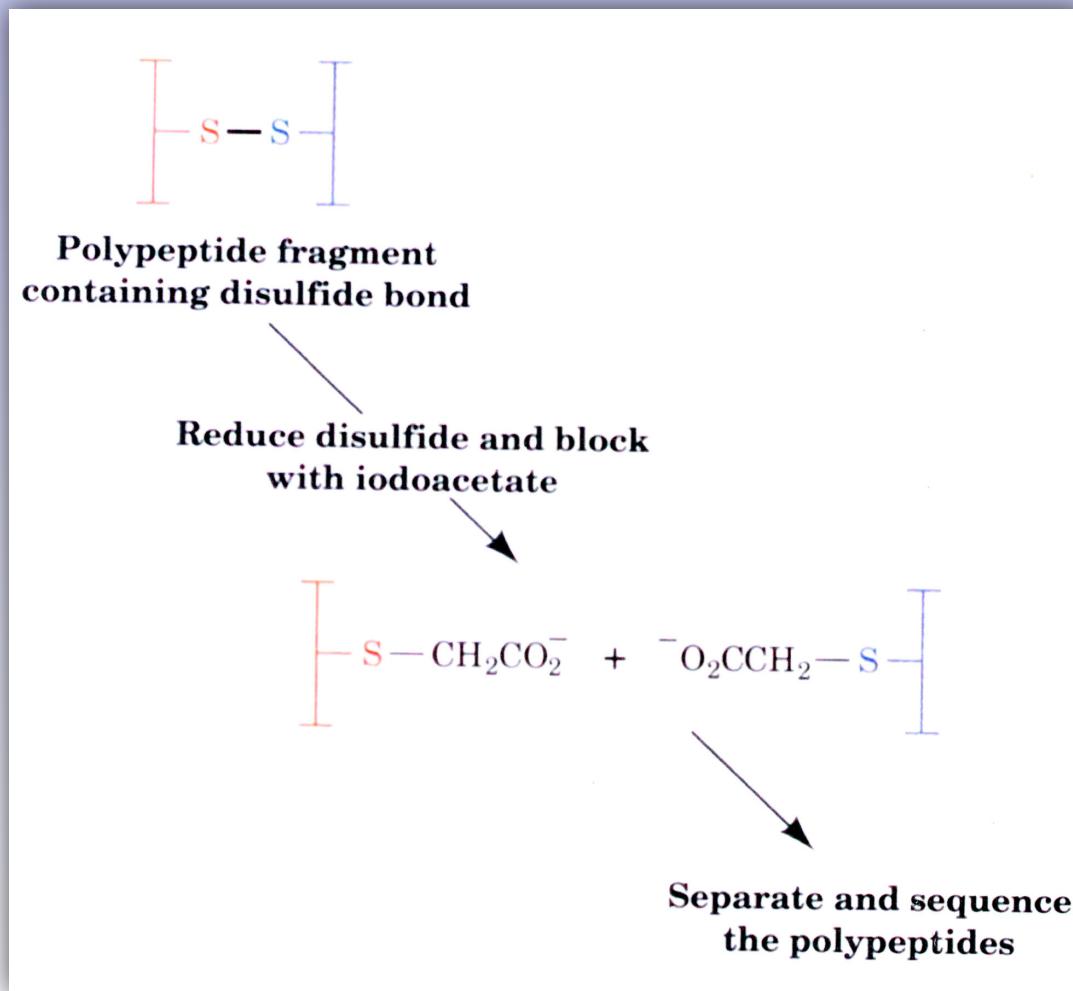
## Sequencing the proteolytic fragments

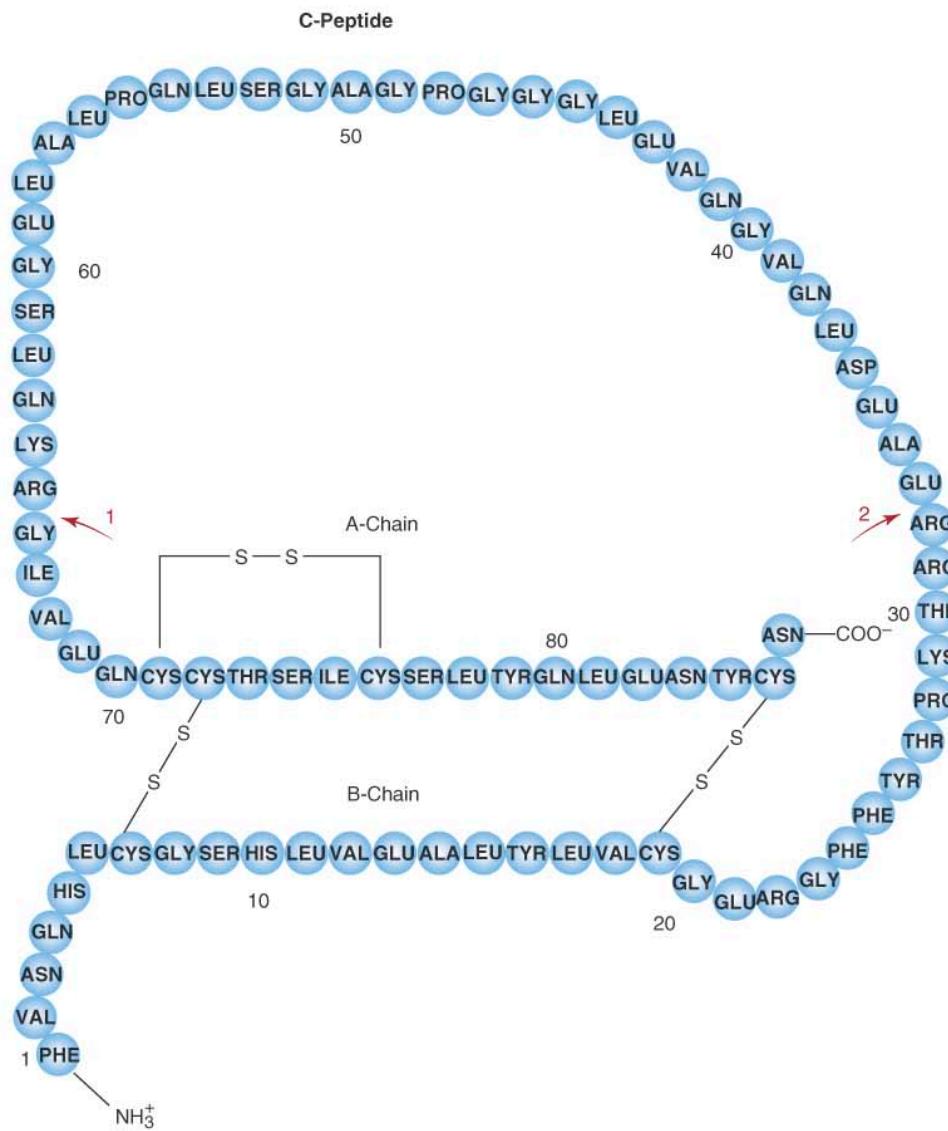
**Edman degradation.**  
 Acid hydrolysis of the PTC polypeptide yields the PTH amino acid (which is identified analytically) and the intact polypeptide minus one amino acid from the N-terminus.



**Primary structure of bovine insulin.** The native molecule is comprised of two separate oligopeptide chains linked at two sites by interchain disulfide bonds (cystine). An intrachain disulfide bond is also present in the native structure.

## Reductive cleavage and alkylation of a disulfide bond

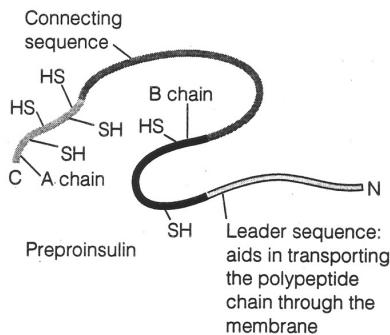




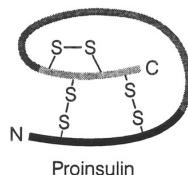
**Figure 6.19. Maturation of human proinsulin.** Redrawn from Bell, G. I., Swain, W. F., Pictet, R., Cordell, B., Goodman, H.M., and Rutter, W. J. *Nature* 282:525, 1979.

**Figure 5.21 Structure of preproinsulin and its conversion to insulin**

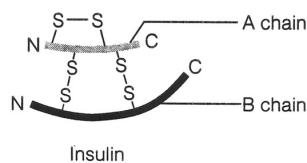
- ① Preproinsulin is synthesized as a random coil on membrane-associated ribosomes



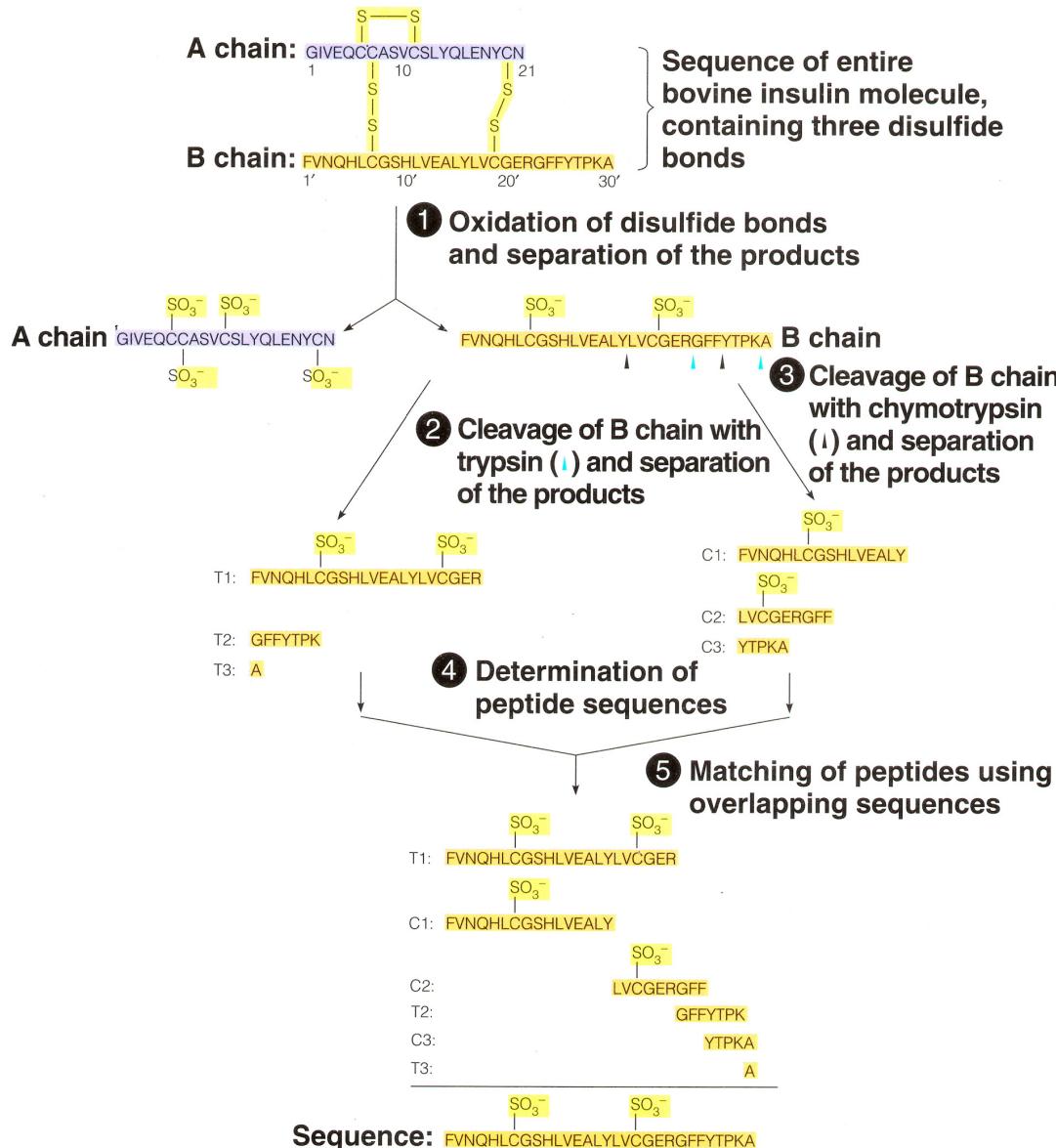
- ② After membrane transport, the leader sequence is cleaved and the resulting proinsulin folds into a stable conformation  
③ Disulfide bonds form

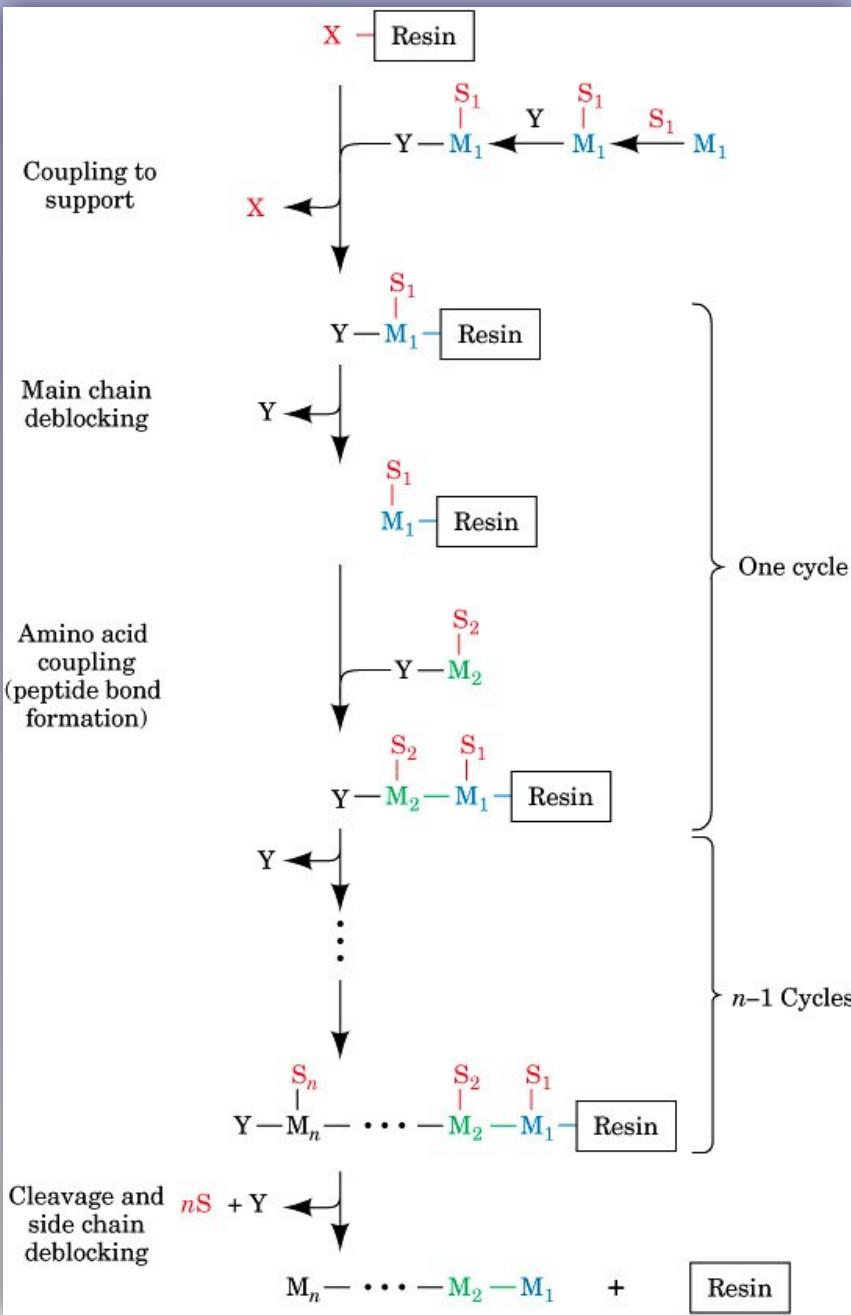


- ④ The connecting sequence is cleaved to form the mature insulin molecule



**Figure 5D.1 Sequencing the  $\beta$  chain of insulin**

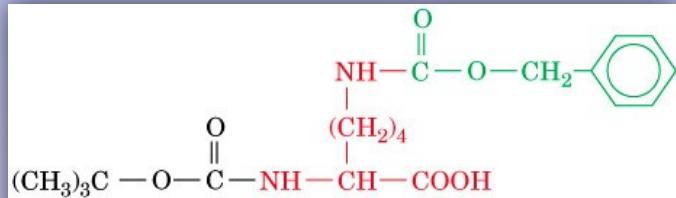




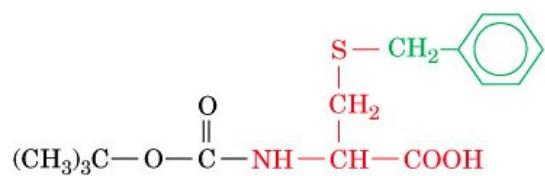
## Solid-phase synthesis of peptides and proteins

Flow diagram for the chemical synthesis of a polypeptide by the **solid phase method (Merrifield synthesis)**

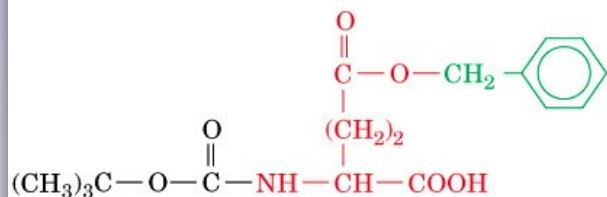
Synthesis direction:  
C-terminus to N-terminus



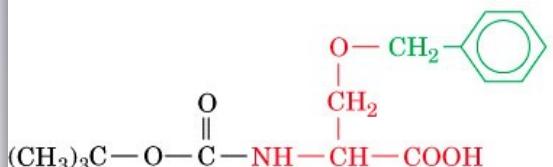
### **Boc, N<sup>ε</sup>-benzyloxycarbonyl-Lys**



### Boc, S-benzyl-Cys



### Boc-Glu, $\gamma$ -Benzyl ester

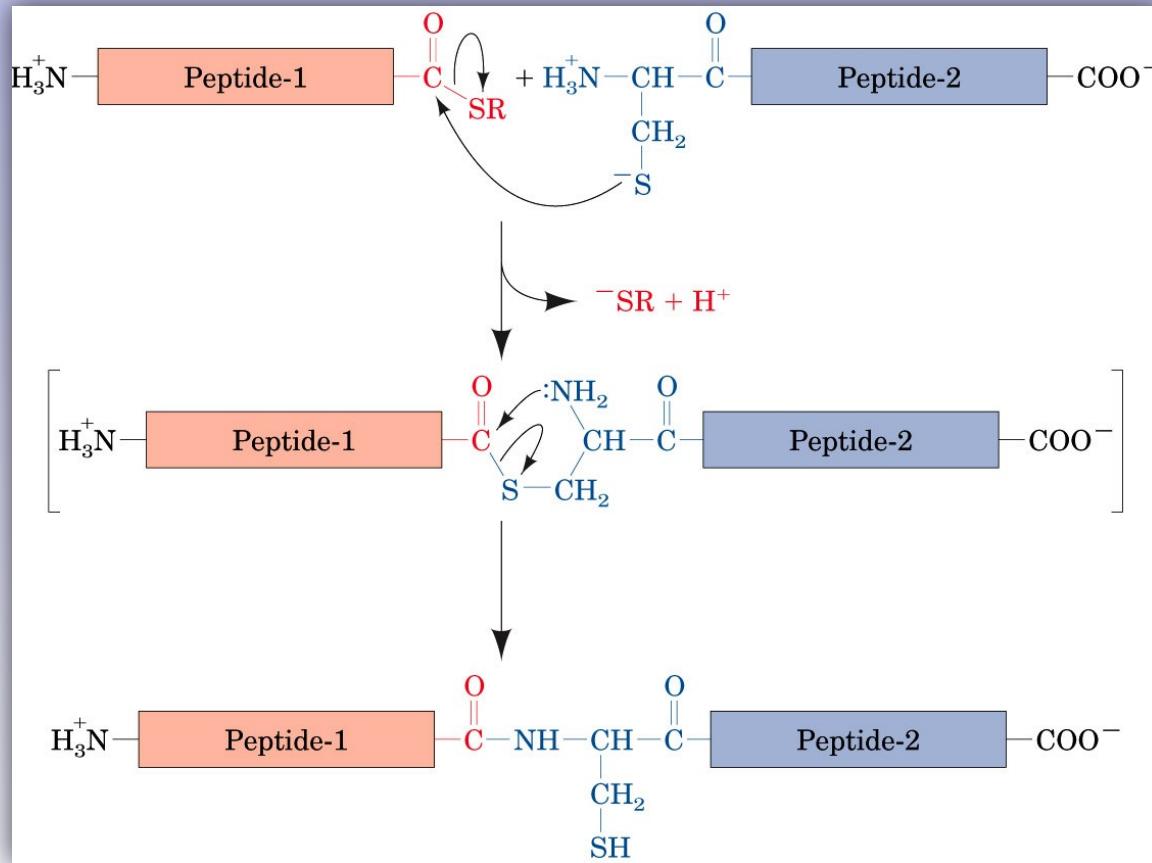


### Boc, O-benzyl-Ser

# Some amino acid derivatives containing benzyl-protected sidechains and BOC-protected $\alpha$ -amino groups used in solid-phase peptide synthesis

**BOC = *t*-butyloxycarbonyl**

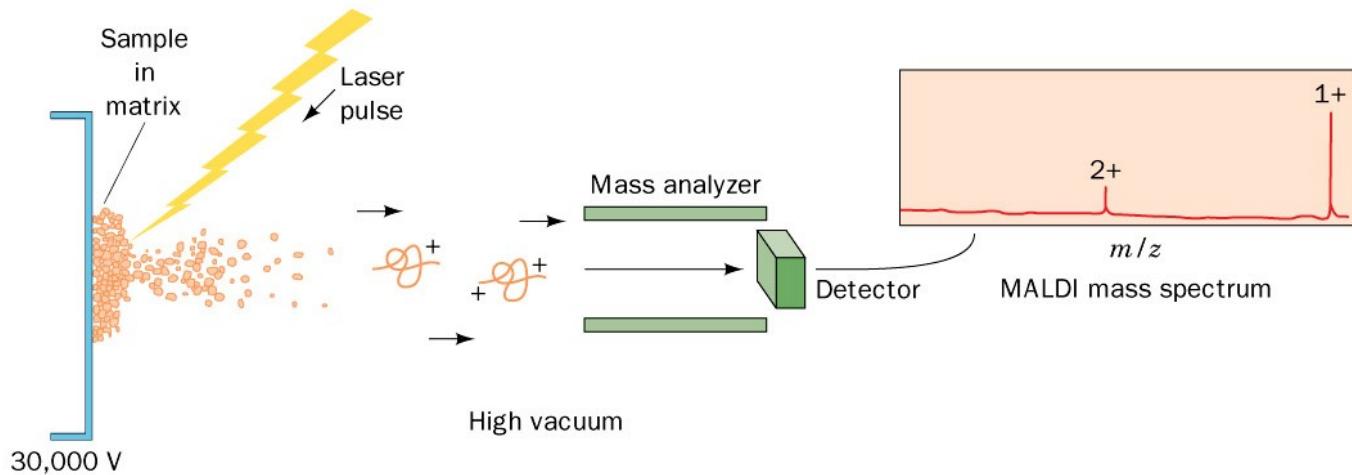
## Linking two peptides together chemically



Connecting two peptides via chemical coupling:  
native chemical ligation

# **Analytical tools for peptide/protein characterization: Mass spectrometry**

(b) Matrix-assisted laser desorption/ionization (MALDI)



**Generation of gas-phase ions required for mass spectrometric analysis of proteins:  
matrix-assisted laser desorption/ionization (MALDI)**

**Bioinformatics:** intersection of biotechnology and computer science; computational tools and methods used to extract useful structural information and relationships from protein and DNA sequence data

- sequence databases
- sequence alignment (homology; pairwise and multiple alignments)
- phylogenetic relationships

Mb	G	L	S	D	G	E	W	Q	L	V	L	N	V	W	G	K	V	E	A	D	I	P	G	H	G	Q	E	V	L	I	R	L	F	K	G	H	P	E	T	L	40
Hb $\alpha$	V	L	S	P	A	D	K	T	N	V	K	A	A	W	G	K	V	G	A	H	A	G	E	Y	G	A	E	A	L	E	R	M	F	L	S	F	P	T	T	K	40
Mb	E	K	F	D	K	F	K	H	L	K	S	E	D	E	M	K	A	S	E	D	L	K	K	H	G	A	T	V	L	T	A	L	G	G	I	L	K	K	K	G	80
Hb $\alpha$	T	Y	F	P	H	F	-	-	-	-	-	D	L	S	H	G	S	A	Q	V	K	G	H	G	K	K	V	A	D	A	L	T	N	A	V	A	H	V	D	74	
Mb	H	H	E	A	E	I	K	P	L	A	Q	S	H	A	T	K	H	K	I	P	V	K	Y	L	E	F	I	S	E	C	I	I	Q	V	L	Q	S	K	H	P	120
Hb $\alpha$	D	M	P	N	A	L	S	A	L	S	D	L	H	A	H	K	L	R	V	D	P	V	N	F	K	L	L	S	H	C	L	L	V	T	L	A	A	H	L	P	114
Mb	G	D	F	G	A	D	A	Q	G	A	M	N	K	A	L	E	L	F	R	K	D	M	A	S	N	Y	K	E	L	G	F	Q	G	153							
Hb $\alpha$	A	E	F	T	P	A	V	H	A	S	L	D	K	F	L	A	S	V	S	T	V	L	T	S	K	Y	R	141													

AS = 365      NAS = 259      % ID = 27.0

Optical alignments of human myoglobin (Mb, 153 residues) and the human hemoglobin  $\alpha$  chain (Hb $\alpha$ , 141 residues)

AS = alignment score; NAS = normalized alignment score;  
%ID = percent identical

### (a) BLAST pairwise alignment

>sp|P38524|HPI2\_ECTVA HIGH POTENTIAL IRON-SULFUR PROTEIN, ISOZYME 2 (HIPPIP 2)  
Length = 71

Score = 50.4 bits (118), Expect = 6e-07  
Identities = 27/69 (39%), Positives = 35/69 (50%), Gaps = 4/69 (5%)

**Query: 1** EPRAEDGHAHDYVNEAADASG--HPRYQEGQLCENCASFGEAVQDGWGRCTHPDFDEVLVKAEGWCSVY 67  
E +ED A + DAS HP Y+EQQ C NC + +A WG C+ F LV A GWC+ +  
**Sbjct: 2** ERLSEDDPAAQALEYRHDASSVQHPAYEEGQTCLNCLLYTDASAQDWGPCS--VFPGKLVSANGWCTAW 68

**(b) FASTA pairwise alignment**

>>SWALL:HPI2 ECTVA P38524 HIGH POTENTIAL IRON-SULFUR PRO (71 aa)

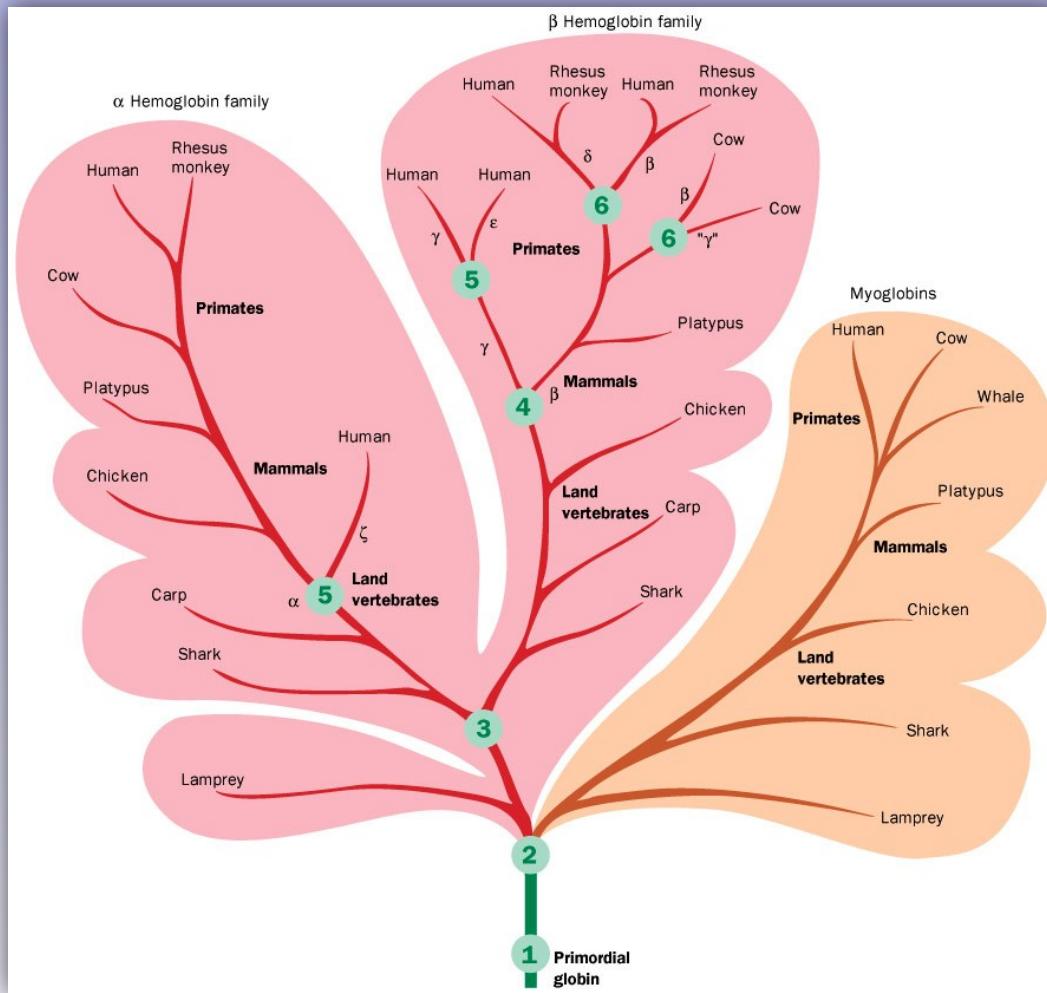
initn: 102 init1: 77 opt: 116 Z-score: 278.0 expect() 4e-08

Smith-Waterman score: 116; 39.130% identity in 69 aa overlap (1-67:2-68)

**(c) CLUSTAL X multiple-sequence alignment**

1	sp P38524 HPI2	----MERLSEDDPAAQALEYRH <ins>DASSVQ</ins> -HPAYE---EGQTCLNCLLYTDASAQDWGPC--SVFPGKLV <ins>SANGWCTAWVAR</ins>
2	sp P38941 HPI1	----AERLDENSPEALALNYKH <ins>DGASVD</ins> -HP SHA---AGQKCINCLLYTDP SATEWG <ins>GC</ins> --AVFPNKLV <ins>NANGWCTAYVARG</ins>
3	sp P00265 HPI5	----APVDEKNPQAVALGYVS <ins>DAAKAD</ins> -KAKYKQFVAGSHCGNCALFQGKATDAVGGC--PLFLAGKQVANK <ins>GWCSAWAKKA</ins>
4	sp P04168 HPI1	-----EPRAEDGH <ins>AHDYVN</ins> EAADASGH <ins>PRYQ</ins> --EGQLCENCAF <ins>WGEAV</ins> QDGWGRCTH <ins>PDFDEV</ins> LVAE <ins>GWCSVYAPAS</ins>
5	sp P04169 HPI2	GLPDGV <ins>EDLPKA</ins> EDDH <ins>AHDYVN</ins> DAADTD-HARFQ---EGQLCENCQFWVDYVN-GWGYCQHPDFTDVLVR <ins>EGWCSVYAPA</ins> -

Examples of peptide *pairwise* and *multiple* sequence alignments:  
BLAST (basic local alignment search tool); FASTA; CLUSTAL



**Phylogenetic tree** of the globin family. Circled branch points represent gene duplications, and unmarked branch points are species divergences.

# **Web addresses for the major protein and DNA sequence data banks**

## ***Data Banks Containing Protein Sequences***

### **ExPASy Molecular Biology Server (SWISS-PROT):**

<http://expasy.ch/>

### **Protein Information Resource (PIR):**

<http://pir.georgetown.edu/>

### **Protein Research Foundation (PRF):**

<http://www.prf.or.jp/en/>

## ***Data Banks Containing Gene Sequences***

### **GenBank:**

<http://www.ncbi.nlm.nih.gov/Genbank/GenbankSearch.html>

### **European Bioinformatics Institute (EBI):**

<http://srs.ebi.ac.uk/>

### **DBGET/LinkDB Integrated Database Retrieval System:**

<http://www.genome.ad.jp/dbget/>