

# TECHNICAL NOTES

## 4-Azido[3,5-<sup>3</sup>H]phenacyl Bromide, a Versatile Bifunctional Reagent for Photoaffinity Radiolabeling. Synthesis of Prostaglandin 4-Azido[3,5-<sup>3</sup>H]phenacyl Esters

Bradley D. Smith,<sup>†</sup> Koji Nakanishi,<sup>\*,†</sup> Kikuko Watanabe,<sup>‡</sup> and Seiji Ito<sup>‡</sup>

Department of Chemistry, Columbia University, New York, New York, 10027, and Osaka Bioscience Institute, 6-2-4 Furuedai, Suita, Osaka 565, Japan. Received August 9, 1990

4-Azido[3,5-<sup>3</sup>H]phenacyl bromide was synthesized in three steps from 4-amino-3,5-diiodoacetophenone and coupled to the prostaglandins PGE<sub>2</sub> and PGD<sub>2</sub> to provide potential photoaffinity compounds.

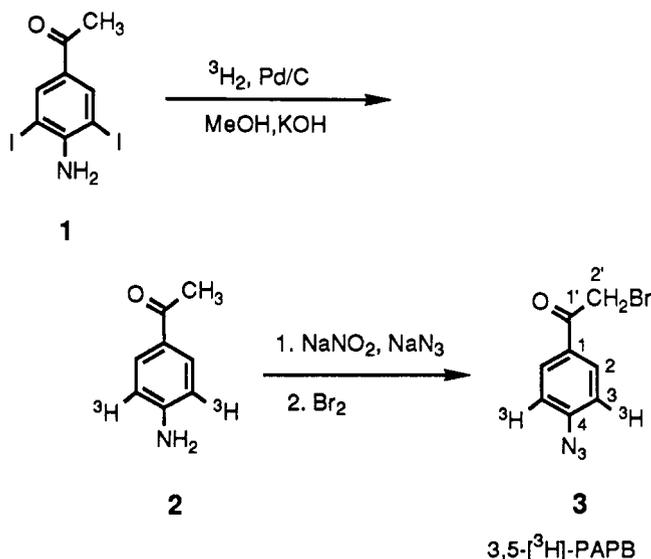
Since its introduction in 1973 (1,2), 4-azidophenacyl bromide (PAPB) has become a useful bifunctional reagent for the modification of both large and small molecules for photolabeling experiments (3). A current literature survey includes more than 25 photoaffinity studies using this compound. Generally, PAPB has been used to selectively alkylate reactive thiol residues in proteins, which have subsequently been used in photo-cross-linking experiments (4, 5). In analogous studies, it has been incorporated into polynucleotides via thiophosphate (6) and thiopyrimidine (7) linkages. Smaller molecules have been modified via attachment to thiol (8), carboxyl (9) and amino (9, 10) moieties to provide photolabile derivatives useful for labeling enzyme and receptor binding sites. In many cases, detection of the photolabeled compounds was achieved by using immunoassay techniques that were specific for each respective system.

We were interested in PAPB as a reagent to modify prostaglandin (PG) compounds for use in photoaffinity experiments (11). We reasoned that a suitable radioactive form of PAPB would provide an efficient way of introducing both the radio- and photolabile groups into the PG molecule. Radiolabeled PAPB has previously been reported

as its 1'-<sup>14</sup>C (2) and 2'-<sup>3</sup>H (12) labeled forms. However, both derivatives were considered unsuitable for our purposes; the specific activity of the <sup>14</sup>C compound was too low, while synthesis of the <sup>3</sup>H compound required specialized handling. Also considered as unsuitable were any potential derivatives labeled with <sup>125</sup>I, since it has been noted that arylazido compounds substituted with iodine sometime result in low incorporation of the photoprobe (13).

We therefore decided to synthesize [3,5-<sup>3</sup>H]PAPB (3), which was achieved by the sequence described in Scheme I. The tritium was introduced by catalytic dehalogenation of 4-amino-3,5-diiodoacetophenone (1; obtained by reaction of 4-aminoacetophenone with 2 equiv of iodine monochloride) using tritium gas and Pd/C catalyst in methanol/KOH.<sup>1</sup> Under these conditions the incorporation of tritium was high (specific activity of 50 Ci/mmol) and no concomitant reduction of the aryl ketone function was observed. Without purification, the tritiated 4-aminoacetophenone (2) was converted to [3,5-<sup>3</sup>H]-PAPB (3) in a simple, high-yielding, two-step process.<sup>2</sup> As

### Scheme I



<sup>1</sup>The following was carried out by Amersham Corp., tritium labeling service (TR3 method): 4-amino-3,5-diiodoacetophenone (40 mg), 10% Pd/C (4 mg), methanol (8 mL), and 1 N KOH aqueous methanol solution (0.50 mL) were hydrogenated at room temperature under 1 atm of tritium gas for 3 h. The catalyst and solvent were removed, and the tritiated product 2 (14 mg, 5 Ci, 50 Ci/mmol) was taken up in ethanol (30 mL).

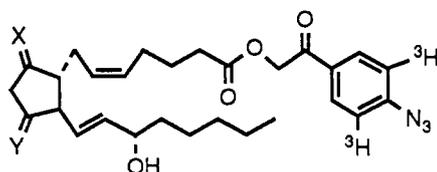
<sup>2</sup>To a 2.5-mL aliquot of the stock ethanol solution of tritiated 2 (500 mCi, 1.2 mg) was added unlabeled 4-aminoacetophenone (10.0 mg) and 5% aqueous H<sub>2</sub>SO<sub>4</sub> (5 mL). The solution was cooled in ice, treated with aqueous NaNO<sub>2</sub> solution (18 mg in 200 μL), and stirred for 20 min before a chilled solution of NaN<sub>3</sub> (24 mg in 200 μL) was added. After stirring for a further 10 min, ether (10 mL) was added, and the phases separated. The organic layer was washed (2 × 3 mL water), dried (MgSO<sub>4</sub>), and evaporated to give crude 4-azido[3,5-<sup>3</sup>H]acetophenone. This residue was taken up in ether (1 mL) and treated with acetic acid (1 drop) and bromine solution (60 μL, 1 N CCl<sub>4</sub> solution). After 40 min, TLC indicated the bromination was complete [*R*<sub>f</sub> for 4-azidoacetophenone and monobromo and dibromo products (eluent, CH<sub>2</sub>Cl<sub>2</sub>) were 0.56, 0.74, 0.82; product ratios were 0.05, 0.90, 0.05, respectively]. Ether (15 mL) and water (3 mL) were added, and the organic layer separated and was dried. Removing of the solvent and purification by flash chromatography (1.5 × 30 cm column packed with 12 cm of silica; eluent, 1:1 CH<sub>2</sub>Cl<sub>2</sub>/hexane) gave [3,5-<sup>3</sup>H]PAPB (3) [8.0 mg (38%), 160 mCi, 4.8 Ci/mmol; UV MeOH, λ<sub>max</sub> 292 nm, ε = 2.08 × 10<sup>4</sup> (2)], which was taken up in toluene (5 mL) and stored at -78 °C.

<sup>†</sup> Columbia University.

<sup>‡</sup> Osaka Bioscience Institute.

a bifunctional reagent, [3,5-<sup>3</sup>H]PAPB has the following advantages: (i) Its synthesis is straightforward and inexpensive, producing large amounts of labeled material of high specific activity. (ii) [3,5-<sup>3</sup>H]PAPB is stable to long-term storage at low temperature and therefore, in principle, one sample can be utilized for a variety of labeling targets. Even unreactive amino and alcohol groups can be attached via linker groups (5). (iii) The presence of the radio- and photolabels on the same ring eliminates the possibility of them becoming separated in any postlabeling digestive workup.

We have used [3,5-<sup>3</sup>H]PAPB to esterify the carboxyl functions of PGE<sub>2</sub> and PGD<sub>2</sub> to produce potential PG photoaffinity compounds 4 and 5, respectively.<sup>3</sup> These PG compounds are currently being used in photolabeling experiments designed to characterize the structural aspects of PG synthase enzymes (14) and PG receptors (15).



4 X = O; Y =  $\alpha$ -OH,  $\beta$ -H

5 X =  $\alpha$ -OH,  $\beta$ -H; Y = O

#### ACKNOWLEDGMENT

This study was supported by NIH 10187. B.D.S. gratefully acknowledges a fellowship from Merck, Sharp and Dohme. We are grateful to Professor O. Hayaishi for discussions and encouragement.

**Note Added in Proof:** The procedures described in footnotes 2 and 3 have been successfully repeated on a 10-fold smaller scale to allow the carrier-free synthesis of [3,5-<sup>3</sup>H]PAPB and prostaglandin ester 4 at a specific activity of 50 Ci/mmol.

#### LITERATURE CITED

- Hixson, S. S., and Hixson, S. H. (1973) Photochemical labeling of yeast alcohol dehydrogenase with an azide analogy of NAD<sup>+</sup>. *Photochem. Photobiol.* 18, 135-8.
- Hixson, S. H., and Hixson, S. S. (1975) *p*-Azidophenacyl bromide, a versatile photolabile bifunctional reagent. Reaction with glyceraldehyde-3-phosphate dehydrogenase. *Biochemistry* 14, 4251-4.
- Bayley, H. (1983) *Photogenerated Reagents in Biochemistry and Molecular Biology*, Elsevier, New York.
- Some examples: First, E. A., and Taylor, S. S. (1988) Subunit interaction sites between the regulatory and catalytic subunits of cAMP-dependent protein kinase. Heterobifunctional cross-linking reagents lead to photodependent and photoindependent cross-linking. *J. Biol. Chem.* 263, 5170-5. Lunn, C. A., and Pigiet, V. P. (1986) Chemical cross-linking of thioredoxin to hybrid membrane fraction in *Escherichia coli*. *J. Biol. Chem.* 261, 832-8. Dupuis, A., and Vignais, P. V. (1985) Photolabeling of mitochondrial F1-ATPase by an azido derivative of the oligomycin-sensitivity conferring protein. *Biochem. Biophys. Res. Commun.* 129, 819-25.
- Erecinska, M. (1980) The use of photoaffinity labels in the study of mitochondrial function. *Ann. N. Y. Acad. Sci.* 346, 444-57.
- Praseuth, D., Perroualt, L., Trung, D., Chassignol, M., Nguyen, T., and Helene, C. (1988) Sequence Specific binding and photocrosslinking of  $\alpha$  and  $\beta$  oligonucleotides to the major groove of DNA via triple helix formation. *Proc. Natl. Acad. Sci. U.S.A.* 85, 1349-53. Hanna, M. M., and Meares, C. F. (1983) Topography of transcription: Path of the leading end of nascent RNA through the *Escherichia coli* transcription complex. *Proc. Natl. Acad. Sci. U.S.A.* 80, 4238-42. Hanna, M. M., and Meares, C. F. (1983) Synthesis of a cleavable dinucleotide photoaffinity probe of ribonucleic acid polymerase: Application to trinucleotide labeling of an *Escherichia coli* transcription complex. *Biochemistry*, 22, 3546-51.
- Some examples: Hanna, M. M., Dissinger, S., Williams, B. D., and Colston, J. E. (1989) Synthesis and characterization of 5-(4-azidophenacyl)thiouridine 5'-triphosphate, a cleavable photo-cross-linking nucleotide analog. *Biochemistry*, 28, 5814-20. Hsu, L. M., Lin, F. L., Nurse, K., and Ofengand, J. (1984) Covalent cross-linking of *Escherichia coli* phenylalanyl-tRNA and valyl-tRNA to the ribosomal A site via photoaffinity probes attached to the 4-thiouridine residue. *J. Mol. Biol.* 172, 57-76.
- Some examples: Kunst, M., Sies, H., and Akerboom, T. P. (1989) *S*-(4-azidophenacyl)[<sup>35</sup>S]-glutathione photoaffinity labeling of rat liver plasma membrane-associated proteins. *Biochem. Biophys. Acta* 982, 15-23. Matthew, M. S., Lewis, R. V., and Barden, R. E. (1986) Photoaffinity labeling of carnitine acetyltransferase with *S*-(*p*-azidophenacyl)thiocarnitine. *Biochem. J.* 237, 533-40.
- Yurchenko, R. I., and Malitskaya, V. P. (1977) Syntheses based on *p*-azido- $\omega$ -haloacetophenones. *Zh. Org. Khim.* 13, 1980-7.
- Deushkin, Y. V., and Kotelevtsev, Y. V. (1982) Photoaffinity modification by azidocytisine of the nicotine acetylcholine receptor from optic ganglia of the squid. *Bioorg. Khim.* 13, 1980-7.
- Michalak, M., Wandler, E. L., Strynadka, K., Lopaschuk, G. L., Njue, W. M., Liu, H.-J., and Olley, P. M. (1990) Photolabeling of the prostaglandin E<sub>2</sub> receptor in cardiac sarcolemmal vesicles. *FEBS Lett.* 265, 117-20.
- Muccino, R. R., and Serico, L. (1978) Alumina catalysed exchange of enolizable hydrogens. *J. Labelled Compd. Radiopharm.* 15, 523-7.
- Watt, D. S., Kawada, K., Leyva, E., and Platz, M. S. (1989) Exploratory photochemistry of iodinated aromatic azides. *Tetrahedron Lett.* 30, 899-902.
- Watanabe, K., Fujii, Y., Nakayama, K., Ohkubo, H., Kuramitsu, S., Kagamiyama, H., Nakanishi, S., and Hayaishi, O. (1988) Structural similarity of bovine lung prostaglandin F synthase to lens  $\epsilon$ -crystallin of the european common frog. *Proc. Natl. Acad. Sci. U.S.A.* 85, 11-5.
- Negishi, M., Ito, S., Tanaka, T., Yokohama, H., Hayashi, H., Katada, T., Ui, M., and Hayaishi, O. (1988) Covalent cross-linking of prostaglandin E receptor from bovine adrenal medulla with a pertussis toxin-insensitive guanine nucleotide-binding protein. *J. Biol. Chem.* 262, 12077-84.

<sup>3</sup>[3,5-<sup>3</sup>H]PAPB (3; 4 mg, 16 mmol, 80 mCi) was treated with a solution of PGD<sub>2</sub> (10 mg, 28 mmol) in THF (0.50 mL) and diisopropylethylamine (5  $\mu$ L, 29 mmol). The solution was stirred overnight at room temperature, the solvent was evaporated and the residue was purified by flash chromatography (1  $\times$  20 cm column packed with 6 cm of silica; eluent, 1:1 ethyl acetate/hexane) to give PGD<sub>2</sub> 4-azido[3,5-<sup>3</sup>H]phenacyl ester 5 [4.8 mg, 45 mCi, 4.8 Ci/mmol]. PGE<sub>2</sub> 4-azido[3,5-<sup>3</sup>H]phenacyl ester 4 was synthesized by using the same procedure except the chromatography eluent was 4:1 ethyl acetate/hexane.